BIOPHYSICS LETTER

Paola Llanos · Mauricio Henriquez · Jasmina Minic Khalil Elmorjani · Didier Marion · Gloria Riquelme Jordi Molgó · Evelyne Benoit

Neuronal and muscular alterations caused by two wheat endosperm proteins, puroindoline-a and alpha1-purothionin, are due to ion pore formation

Received: 5 August 2003 / Accepted: 19 August 2003 / Published online: 15 October 2003 © EBSA 2003

Abstract Using the patch-clamp technique it was found that the toxicity of the two wheat endosperm proteins puroindoline-a and alpha1-purothionin probably results from the dissipation of ion concentration gradients essential for the maintenance of cellular homeostasis.

Keywords Giant liposomes · Mouse neuromuscular junction · Neuroblastoma cells · Pore formation · Wheat endosperm proteins

Abbreviations PIN-a puroindoline-a · PTH alpha1purothionin

Introduction

Puroindoline-a (PIN-a) and alpha1-purothionin (PTH), two basic cysteine-rich proteins isolated from the wheat endosperm of Triticum aestivum sp., have been suggested to play a role in plant defence mechanisms against phytopathogenic organisms (Blochet et al. 1993; Bohlmann 1994; Dubreil et al. 1998). The two proteins were shown to have a marked toxicity to vertebrate cells, as revealed by studying their effects on the morphological

Presented at the Biophysical Society Meeting on "Ion Channels—from structure to disease" held in May 2003, Rennes, France

J. Minic · J. Molgó · E. Benoit (⋈) Laboratoire de Neurobiologie Cellulaire et Moléculaire, UPR 9040, Institut Fédératif de Neurobiologie Alfred Fessard, CNRS, bât. 32–33, 91198 Gif sur Yvette Cedex, France E-mail: benoit@nbcm.cnrs-gif.fr

P. Llanos · M. Henriquez · G. Riquelme Instituto de Ciencias Biomédicas, Facultad de Medicina, Universidad de Chile, Independencia No. 1027, Casilla 70005, Santiago 6530499, Chile

K. Elmorjani · D. Marion Laboratoire de Biochimie et Technologie des Protéines, INRA, B.P. 1627, 44316, Nantes Cedex 03, France

Fax: +33-1-69829466

and functional properties of neuroblastoma NG108-15 cells, frog myelinated axons, C2 myotubes and mammalian neuromuscular junctions, using confocal laser scanning microscopy and conventional electrophysiology (Mattei et al. 1998; Benoit et al. 2001). In order to obtain insight into the mechanisms involved in the cellular toxicity of PIN-a and PTH, the proteins were incorporated into asolectin giant liposomes (Riquelme et al. 1990) and their pore-forming ability was investigated using the patch-clamp technique in excised patch configuration.

Results

Current recordings from liposomes containing PIN-a revealed that the protein forms single channels (Fig. 1A) with a unitary conductance of about 15 pS between -80 and 80 mV. The reversal potential of the current recorded in response to potential-ramps was shifted from 0 to -24 mV when the bath concentration of NaCl was increased from 140 to 440 mM, and from 0 to -9 mV when external NaCl was replaced by KCl. This indicates that PIN-a forms cationic channels whose selectivity is $K^+ > Na^+ > > Cl^-$. Current recordings from liposomes containing PTH revealed that this protein also forms single channels (Fig. 1B, C) with a unitary conductance at -100 mV of about 35 pS in 2/3 of the recordings and about 100 pS in 1/3 of the recordings. The ionic selectivity was $Na^+ \approx K^+ > > Cl^-$ for the low conductance channels and $Cl^- > Na^+$ for the large conductance channels, indicating that these channels correspond to cationic and anionic channels, respectively.

Conclusion

It is concluded that the toxicity of the two wheat endosperm proteins, PIN-a and PTH, to fungi, yeast, bacteria and vertebrate cells (Blochet et al. 1993; Bohlmann 1994; Dubreil et al. 1998; Mattei et al. 1998;

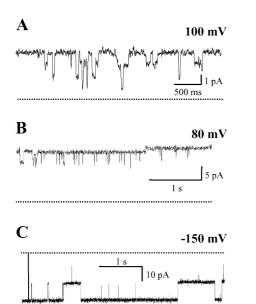


Fig. 1 Ion-channel activities exhibited by giant liposomes containing either PIN-a (A) or PTH (B) and (C). The single-current traces were recorded at the indicated holding potential. The zero current level is indicated by the *dotted lines*

Benoit et al. 2001) is likely to result from the dissipation of ion concentration gradients essential for the maintenance of cellular homeostasis.

Acknowledgements The authors are grateful to M. Malo and B. Rouzaire-Dubois for providing the neuroblastoma (NG108-15) cell cultures used in this study.

References

Benoit E, Elmorjani K, Molgó J, Marion D, Minic J (2001) Actions neuronales et musculaires de deux polypeptides de l'albumen de Blé. In: Bon C, Goudey-Perrière F, Poulain B, Puiseux-Dao S (eds), Rencontres en toxinologie: Explorer, exploiter les toxines et maîtriser les organismes producteurs. Elsevier, Paris, pp 79–80

Blochet JE, Chevalier C, Forest E, Pebay-Peyroula E, Gautier MF, Joudrier P, Pezolet M, Marion D (1993) Complete amino acid sequence of puroindoline, a new basic and cystine-rich protein with a unique tryptophan-rich domain, isolated from wheat endosperm by Triton X-114 phase partitioning. FEBS Lett 329:336–340

Bohlmann H (1994) The role of thionins in plant protection. Crit Rev Plant Sci 13:1–16

Dubreil L, Gaborit T, Bouchet B, Gallant DJ, Broekaert W, Quillien L, Marion D (1998) Spatial and temporal distribution of the major isoforms of puroindolines (puroindoline-a and puroindoline-b) and non specific lipid transfer protein (ns-LTPe1) of *Triticum aestivum* seeds. Relationships with their in vitro antifungal properties. Plant Sci 138:121–135

Mattei C, Elmorjani K, Molgó J, Marion D, Benoit E (1998) The wheat proteins puroindoline-a and alpha1-purothionin induce nodal swelling in myelinated axons. Neuroreport 9:3803–3807

Riquelme G, Lopez E, Garcia-Segura LM, Ferragut JA, Gonzalez-Ros JM (1990) Giant liposomes: a model system in which to obtain patch-clamp recordings of ionic channels. Biochemistry 29:11215–11222